assumes the very rapid establishment of equilibrium between molecules and ion pairs in the coupled equilibria, molecules \rightleftharpoons ion pairs \rightleftharpoons free ions, and we therefore conclude that in our systems the formation of the loose ion pair from the tight ion pair is not rate determining. This agrees with the conclusions reached from our kinetic studies8 on NiLCl₂.

Hideo Hirohara, Kenneth J. Ivin, John J. McGarvev*

Department of Chemistry, The Queen's University of Belfast Belfast BT9 5AG, Northern Ireland Received January 3, 1974

²³Na Quadrupole Coupling Constants in Different Coordination Shells from ²³Na and ¹³C Fourier **Transform Nuclear Magnetic Resonance** Measurements on Sodium Cryptates

Sir:

Ionic solvation has been investigated by many research groups using most of the available electrochemical and spectroscopic methods, including nuclear magnetic resonance spectroscopy.¹ Among metal cations, much work has been done on the sodium cation by direct observation on the ²³Na nmr chemical shifts and quadrupolar broadening effects.²⁻¹³

Knowledge of ²³Na quadrupolar coupling constants, $\chi_{\rm Na} = (e^2 q Q/h) (eQ = {}^{23}{\rm Na}$ nuclear quadrupole moment), and electric field gradients, eq in various coordination states, would be of much interest to the understanding of the nature of cation solvation, but such determinations are hampered by the uncertainties in solvation number¹⁴ and the fluctuation of the coordination sphere.

Stable sodium complexes with well-defined coordination shells provide a unique way to study the solvated cation, as shown by recent ²³Na studies¹⁵⁻¹⁷ of Na+ complexes of natural¹⁵ and synthetic¹⁸ macrocyclic ligands.

(1) J. F. Hinton and E. S. Amis, Chem. Rev., 67, 367 (1967); U. Mayer and V. Gutman, Struct. Bonding (Berlin), 12, 113 (1972); C. Deverell, Progr. Nucl. Magn. Resonance Spectrosc., 4, 235 (1969).
(2) O. Jardetzky and J. E. Wertz, J. Amer. Chem. Soc., 82, 318 (1960).

(3) G. A. Rechnitz and S. B. Zamochnick, J. Amer. Chem. Soc.,

- 86, 2953 (1964). (4) (a) C. Deverell and R. E. Richards, *Mol. Phys.*, **10**, 551 (1966); (b) C. Deverell, *ibid.*, **16**, 491 (1969); (c) C. Hall, R. E. Richards, G. N.
- Schulz, and R. R. Sharp, ibid., 16, 529 (1969).
- (5) M. Eisenstadt and H. L. Friedman, J. Chem. Phys., 46, 2182 (1967).
- (6) V. S. Griffiths and G. Socrates, J. Mol. Spectrosc., 27, 358 (1968).
- (7) E. G. Bloor and R. G. Kidd, Can. J. Chem., 46, 3425 (1968).
- (8) V. I. Ionov, R. K. Mazitov, and I. I. Evdokimov, Zh. Strukt. Khim., 10, 208 (1969).
- (9) T. L. James and J. H. Noggle, Proc. Nat. Acad. Sci. U. S., 62, 644 (1969); J. Amer. Chem. Soc., 91, 3424 (1969).
- (10) R. H. Erlich, E. Roach, and A. I. Popov, J. Amer. Chem. Soc.,
- 92, 4989 (1970); R. H. Erlich and A. I. Popov, *ibid.*, 93, 5620 (1971); M. Herlem and A. I. Popov, *ibid.*, 94, 1431 (1972); R. H. Erlich, M. S.

- (11) R. D. Green and J. S. Martin, Can. J. Chem., 50, 3935 (1972).
 (11) R. D. Green and J. S. Martin, Can. J. Chem., 50, 3935 (1972).
 (12) G. J. Templeman and A. L. Van Geet, J. Amer. Chem. Soc., 94, 5578 (1972);
 (12) G. J. Campleman and A. L. Van Geet, J. Amer. Chem. Soc., 94, 5578 (1972);
- (13) G. W. Canters, J. Amer. Chem. Soc., 94, 5230 (1972).
 (14) E. S. Amis and J. F. Hinton, Chem. Rev., 71, 627 (1971).
- (15) D. H. Haynes, B. C. Pressman, and A. Kowalsky, Biochemistry,
- 10,852 (1971). (16) A. M. Grotens, J. Smid, and E. De Boer, Chem. Commun., 759
- (1971) (17) E. Shchori, J. Jagur-Grodzinski, Z. Luz, and M. Shporer, J.
- Amer. Chem. Soc., 93, 7133 (1971); E. Shchori, J. Jagur-Grodzinski, and M. Shporer, *ibid.*, 95, 3842 (1973).
- (18) C. J. Pedersen and H. K. Frensdorff, Angew. Chem., 84, 16 (1972); Angew. Chem., Int. Ed. Engl., 11, 16 (1972).

Macrobicyclic ligands¹⁹ forming cryptate-type cation inclusion complexes, 20 in which the coordination shell entirely surrounds the cation, should be well suited for such studies since these complexes are much more stable²¹ than the macrocyclic ones and cation exchange is slower (see below).

We present here the results of a study of ²³Na nuclear quadrupolar coupling constants in four "frozen" coordination shells containing as binding sites two nitrogens and four, five, or six oxygens or four oxygens and two sulfurs, *i.e.*, the sodium cryptates of ligands 1, 2, 3, and 4, respectively. Assuming zero asymmetry parameter, the quadrupolar relaxation time T_q of the ²³Na nucleus is given by eq 1^{22} (nuclear spin of ²³Na =

$$T_{\rm q}^{-1} = \pi \Delta \nu = (2\pi^2/5)(e^2 q Q/h)^2 \tau_{\rm c}$$
(1)

 $^{3}/_{2}$). χ_{Na} may be calculated from the line width $\Delta \nu$ of the ²³Na resonance if the rotational correlation time, $\tau_{\rm c}$, of the motions of the sodium complex is known. Since τ_{c} values estimated in the usual way from the molecular radius and Debye's relation may be in error (generally too long) by one order of magnitude,²³ we employed an indirect double nuclear spin probe method.^{24,25}

Using the dipole-dipole relaxation equation (2) one

$$T_{1}^{-1}({}^{13}\text{C}) = n^{2}\hbar^{2} \gamma_{\text{H}}^{2} \gamma_{\text{C}}^{2} r_{\text{CH}}^{-6} \tau_{\text{o}} = 4.4046 \times 10^{10} \tau_{\text{o}} \quad (2)$$

$$(n = 2 \text{ for a CH}_{2} \text{ group}; r_{\text{CH}} = 1.085 \pm 0.005 \text{ Å})$$

may obtain τ_c from the ¹³C relaxation times T_1 (¹³C) of the CH₂ carbons of the sodium cryptates. Then, making the reasonable assumption that, in the quite rigid complexes, τ_e (from ¹³C data) also represents the reorientational motions which modulate the ²³Na quadrupolar interaction, one may introduce τ_c in eq 1 and calculate χ_{Na} from $\Delta \nu$. The ¹³C nmr spectra and relaxation times and the 23Na spectra have been measured by the Fourier transform technique. Spectral data and results calculated with eq 1 and 2 are given in Table I.²⁶

The correlation times of the four cryptates (Table I) are about a factor of 5 shorter than those which are cal-

(19) B. Dietrich, J. M. Lehn, J. P. Sauvage, and J. Blanzat, Tetrahedron, 29, 1629 (1973); B. Dietrich, J. M. Lehn, and J. P. Sauvage, Chem. Commun., 1055 (1970).

(20) B. Dietrich, J. M. Lehn, and J. P. Sauvage, Tetrahedron, 29, 1647 (1973).

(21) J. M. Lehn and J. P. Sauvage, Chem. Commun., 440 (1970), and unpublished results; J. M. Lehn, Struct. Bonding (Berlin), 16, 1 (1973).

(22) A. Abragam, "The Principles of Nuclear Magnetism," Clarendon Press, Oxford, 1961, Chapter 8.

(23) W. B. Moniz and H. D. Gutowsky, J. Chem. Phys., 38, 1155 (1962).

(24) J. M. Lehn and J. P. Kintzinger in "Nitrogen N.M.R.," M. Witanowski and G. A. Webb, Ed., Plenum Press, London, 1973, Chapter 3, and references therein

(25) J. P. Behr and J. M. Lehn, Biochem. Biophys. Res. Commun., 49, 1573 (1972); J. R. Lyerla, Jr., D. M. Grant, and R. K. Harris, J. Phys. Chem., 75, 585 (1971).

(26) The solutions contained stoichiometric amounts of a given ligand and NaCl (0.25 M each) in methanol-D₂O 95/5. Since the stability constants are very high in this medium $(>10^\circ)$,²¹ the solution con-tains only the sodium cryptates. The spectra have been measured at 25.1 MHz for both ¹³C (with ²D lock) and ²³Na (without field-frequency lock and by decreasing slightly the magnetic field). The ¹³C relaxation times have been obtained using the inversion-recovery method. The short relaxation times measured (about 1 sec), the size of the molecules and the presence of two protons on each carbon justify the use of eq 2. Indeed the relaxation of these carbon nuclei may be expected to occur almost exclusively via the dipole-dipole mechanism (for a discussion see, for example, G. C. Levy, Accounts Chem. Res., 6, 161 (1973)).

Table I. ¹³C and ²³Na Nmr Spectral Data, Calculated Correlation Times, and ²³Na Quadrupolar Coupling Constants for Sodium Cryptates of Ligands 1–4°

| Cryptate | $T_1({}^{13}C),^b \sec(\pm 15\%)$ | | δ(²³ Na), ^e ppm (±2%) | ∆µ(23Na),¢ Hz | $T_{q}({}^{23}Na),^{d}msec (\pm 5\%)$ | $\chi_{Na},^{d}$ MHz (±10%) |
|-------------------------------------|-----------------------------------|------|---|---------------|---------------------------------------|--------------------------------|
| [1,Na ⁺]Cl ⁻ | 1.05 | 21.5 | +11.15 | 132 ± 3 | 2.4 | 2.20 |
| [2,Na ⁺]Cl ⁻ | 1.25 | 18.0 | -4.25 | 46 ± 2 | 6.9 | 1.43 |
| [3,Na ⁺]Cl ⁻ | 1.00 | 22.5 | -11.40 | 29 ± 1 | 11.0 | 1.01 |
| [4,Na ⁺]Cl ⁻ | 1.10 | 20.5 | -6.20 | 49 ± 2 | 6.5 | 1.38 |

^a See ref 26 for experimental details. ^b ¹³C relaxation times T_1 and correlation times τ_0 calculated according to eq 2. The values given are averages over all carbons in the molecules. The spread of T_1 values is about $\pm 10\%$. In the case of τ_0 an error of ± 0.009 Å in r_{CH} leads to an error of $\pm 5\%$ in τ_0 . τ_0 is given in picoseconds; 1 psec = 10^{-12} sec. ^c δ , shift of ²³Na resonance downfield (<0) from 0.25 *M* NaCl in water (8 Hz line width). $\Delta \nu$, full line width at half-height of ²³Na resonance. A 0.25 *M* NaCl solution in methanol-water has $\delta - 2.70$ ppm and $\Delta \nu = 19.5$ Hz. ^d ²³Na nuclear quadrupolar relaxation time, T_q , and coupling constant, $\chi_{Na} = (e^2 q Q/h)$, calculated using eq 1 and $\tau_0(^{13}C)$ values. T_q is obtained from the full line width $\Delta \nu$; the inhomogeneity contribution to $\Delta \nu$ is probably less than 1-2 Hz leading to an error (over estimation) in χ_{Na} of about 3% or less.

culated from the Debye relation ($\tau_{\rm c} \sim 110$ psec for a molecular radius of about 5 Å).

The ²³Na chemical shifts show an interesting trend. Whereas oxygen containing organic solvents shift the ²³Na resonance downfield, amines give an upfield shift.^{7,10} Indeed the ²³Na resonance is upfield from the reference when the ligand shell contains four oxygens and two nitrogens and shifts downfield as the number of oxygens increases from four to six. Sulfur sites lead to a small downfield shift (Table I).



The ²³Na nuclear quadrupolar coupling constants decrease when the number of oxygens in the ligand shell increases. The electric field symmetry increases as more and more oxygen sites are disposed around the cation. The replacement of two oxygens in 3 by sulfur in ligand 4 causes about the same increase in field gradient as the removal of one oxygen (in 2). The present values of χ_{Na} (Table I) are by a factor of 2–3 higher than those which would be obtained by approximating τ_c with the Debye relation. A linear relation has been proposed^{4b} between χ_{Na} and the paramagnetic shielding term which dominates^{4.7} the ²³Na chemical shifts. Indeed the plot of χ_{Na} against $\delta(Na)$ (Table I) gives a straight line within experimental accuracy.

Finally, when excess NaCl is added to the cryptate solutions two separate ²³Na signals (free and complexed Na⁺) are observed, showing that cation exchange is slow at 35°. In the case of [3, Na⁺] coalescence occurs at about +58° giving a free energy of activation ΔG^{\pm}_{58} of about 15.4 kcal/mol in agreement with previous proton resonance work²⁷ and with a recent ²³Na nmr study of cation exchange rates in the same compound in ethylenediamine.²⁸ In [1,Na⁺] and [2,Na⁺] one still observes separate ²³Na resonances at +58° ($\Delta G^{\pm} > 16$ kcal/mol).²⁹

(28) J. M. Ceraso and J. L. Dye, J. Amer. Chem. Soc., 95, 4432 (1973).
(29) The cation exchange rates in cryptates may be obtained most conveniently from variable temperature ¹³C nmr spectra of 1/1 cryptate-

conveniently from variable temperature ¹³C nmr spectra of 1/1 cryptateligand mixtures (forthcoming report). Double probe ¹³C, ²³Na nmr studies may provide detailed information on the electric effects produced by ligand structure, binding sites, ion pairing, and medium¹⁵ as well as on molecular tumbling. Complexes of other metal cations may also be investigated (lithium and cesium for instance).³⁰

(30) The use of nitrogen-14 as the quadrupolar nucleus allows similar studies to be performed on nitrogen-containing substances of chemical or biological interest.^{24,26}

(31) ĚRA No. 265 from CNRS.

J. P. Kintzinger, J. M. Lehn* Institut de Chimie, Université Louis Pasteur 67-Strasbourg, France³¹ Received October 25, 1973

A Linear Relationship between Substituted Pyridine Lone Pair Vertical Ionization Potentials and pK_a

Sir:

A recent communication¹ on the gas-phase proton affinities, PA, of five 4-substituted pyridines pointed out the importance of the question whether solution phase substituent constants apply in the gas phase.

PA's for a series of bases, B:, will be linear functions of vertical lone pair ionization potentials, IP, provided that the rehybridization energies of B_{ν} ,⁺ and homolytic B:H⁺ bond dissociation energies are constant or are themselves linear functions of IP. If, in addition, the differences in solvation energies between bases, B:, and conjugate acids, BH⁺, are linear functions of IP, a linear relationship should also exist between pK_a and B: lone pair IP. These restrictions preclude the observation of any simple relationship between IP(B:) and pK_a for dissimilar bases, and until now a satisfactory linear pK_a -IP(B:) relationship has not been demonstrated for Brønsted bases.

The photoelectron spectra of ten substituted pyridines have been measured and combined with reported spectra for seven chloropyridines,² methylpyridines,³ and fluoropyridines⁴ from the literature. Spectra were determined on a Perkin-Elmer Model 16 pe spectrometer and calibrated with argon (15.76 eV). To ensure valid

⁽²⁷⁾ J. M. Lehn, J. P. Sauvage, and B. Dietrich, J. Amer. Chem. Soc., 92, 2916 (1970).

M. Taagepera, W. G. Henderson, R. T. C. Brownlee, J. L. Beauchamp, D. Holtz, and R. W. Taft, *J. Amer. Chem. Soc.*, 94, 1369 (1972).
 D. W. Turner, C. Baker, A. D. Baker, and C. R. Brundle, "Mo-

⁽²⁾ D. W. Turner, C. Baker, A. D. Baker, and C. R. Brundle, "Molecular Photoelectron Spectroscopy," Wiley-Interscience, New York, N. Y., 1970.

 ⁽³⁾ E. Heilbronner, V. Hornung, F. Pinkerton, and S. F. Thanes, Helv. Chim. Acta, 55, 289 (1972).

⁽⁴⁾ G. H. King, J. N. Murrel, and R. J. Suffolk, J. Chem. Soc., Dalton Trans., 564 (1972).